

Fig. 2. CPK molecular model of the polypeptide forming a possible receptor site for 5HT shown bound. Aminoacids 107–112 are left out. They form a simple β -turn.

δ-hydrocarbon: ring hydrocarbons to Methyl-Arg, 2. Polar: $-{\rm NH_3}+$ to Gln O: hydroxyl O from Arg $={\rm NH_2}^+$ and ring NH to Gln (102) O. Note that the Gln-Arg carbon-carbon bond (120–221) is twisted by a break in the β -conformation. The β CO = NH hydrogen bond is replaced by a hydrogen bond from Ser (114) OH to peptide O (Pro – 122) and possibly a second from Arg (112) to the next peptide O (123). This twist is necessary for the Arg-5HT interaction described. Gln (120) binds by a hydrogen bond to the carbonyl O of Ala (117).

The complex (Trp-Gln-Arg) somewhat resembles in size, form and charge distribution the molecule of strychnine. This may explain the convulsant activity of the basic protein when injected intraventricularly¹. The function of 5HT in this position might be to maintain this segment of the protein molecule in its β -conformation or to disrupt it by a charge transfer operation (to Trp). This hypothesis can be tested by direct physical investigations using NMR, ORD and CD techniques of the conformation of these polypeptides⁴.

Zusammenfassung. Es besteht die Annahme (CARNEGIE 1970), dass das basische Gehirnprotein als Rezeptor für Serotonin wirkt, und es wird der Vorschlag für molekularbiologische Reaktionen von Serotonin-Rezeptoren gemacht.

J.R. Smythies⁵, F. Benington and R.D. Morin

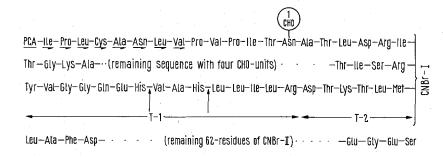
Department of Psychiatry, Universities of Edinburgh and Alabama, and Neuroscience Program, University of Alabama Medical Center, 1919 Seventh Avenue, Birmingham (Alabama 35233, USA), 5 July 1971

- ⁴ F. C. WESTALL, A. B. ROBINSON, J. CACCAIN, J. JACKSON and E. H. EYLAR, Nature. Lond. 229, 22 (1971).
- 5 Acknowledgments. We are most grateful to Dr. D. URRY for advice on protein conformation, to Dr. P. R. Carnegue for helpful discussions and the Ealing Corporation for the generous loan of molecular models.

The Linear Amino Acid Sequence of-α1Acid Glycoprotein

Although α_1 -acid glycoprotein (orosomucoid), a human plasma globulin (for review see ref. 1), is probably the most extensively studied protein with regard to the structure 2 and biosynthesis 3 of its carbohydrate moiety, it is only very recent that significant information concerning its amino acid sequence has been reported 4,5. The complete

elucidation of its primary structure has been hampered by the inability of investigators to establish the aminoterminal sequence, since digestion of α_1 -acid glycoprotein by specific (chymotrypsin and trypsin) and nonspecific (pronase) proteases does not yield overlapping peptides, but affords the same pyrrolidone carboxylic acid, PCA-



The linear amino acid sequence of α_1 -acid glycoprotein. PCA (—) was cleaved specifically with pyrrolidonecarboxylyl peptidase. The amino-terminal sequence of the enzymatically modified CNBr-I was then established by the direct Edman degradation technique (—). The sequence of the carboxyl-terminus of CNBr-I is indicated by the 2 peptides T-1 and T-2. The cleavage of T-1 effected by N-bromosuc cinimide is signified by the 2 arrows (\uparrow).

containing tetrapeptide, PCA-Ile-Pro-Leu. As will be shown in the present paper, this problem could be resolved by cleavage of PCA from the amino-terminal cyanogen bromide-fragment of this glycoprotein (CNBr-I)⁴ by the highly specific enzyme, pyrrolidonecarboxylyl peptidase, and subsequent stepwise chemical degradation of the enzymatically modified CNBr-I. Moreover, the previously unknown peptide sequence that links CNBr-I to the carboxyl-terminal CNBr-fragment (CNBr-II)⁵ of the protein, could also be elucidated.

For the present study α₁-acid glycoprotein was isolated from pooled normal human plasma⁶ and demonstrated to be homogenous as judged by several criteria of purity ^{1,7}. This globulin possesses a single polypeptide chain⁷ to which 5 heteropolysaccharide units⁸, accounting for 40% of the protein by weight, are attached.

After cleavage of this glycoprotein with CNBr and subsequent reduction and alkylation, CNBr-I was isolated by Sephadex G-100 chromatography. For the removal of the amino-terminal pyrrolidone carboxylic acid residue, CNBr-I was treated with pyrrolidonecarboxylyl peptidase⁹. Another aliquot of CNBr-I was trifluoroacetylated 10 and digested with TPCK-trypsin. The resulting digest was passed through a Sephadex G-50 column in order to achieve an initial fractionation into a glycopeptide and a peptide fraction. The carbohydrate-free peptides, after removal of their trifluoroacetyl residues, were separated from each other and purified by conventional procedures 11. One of the resulting peptides (T) that distinguished itself by its content of homoserine, indicating that it was derived from the carboxyl-terminus of CNBr-I, was selected for further investigation. Homogeneity of this peptide was established by high-voltage electrophoresis at pH 1.8 and 6.4, and by amino-terminal amino acid analysis. The amino acid sequence was established by the direct 12 and indirect 13 Edman procedures and by the analysis of carboxypeptidases A and B digests. Chemical cleavage at the histidine linkages was achieved with N-bromosuccinimide (NBS) 11.

The results of the present study reveal the following findings: 1. The specific enzymatic degradation of CNBr-I provides conclusive evidence that PCA represents the amino-terminus of this fragment, thus providing additional and corroborative evidence that PCA indeed is the amino-terminal residue of α_1 -acid glycoprotein as reported earlier 5 . 2. After cleavage of PCA the enzymatically modified CNBr-I was directly accessible to Edman degradation. Eight steps (direct Edman procedure) afforded the sequence which overlapped by 5 residues with the previously established sequence of the 22-amino acid glycopeptide derived from CNBr-I 4 (Figure); and 3. One carbohydrate unit is in close proximity to the amino-terminus of the protein being attached to the 15 th residue (Figure).

For the elucidation of the carboxyl-terminal region of CNBr-I, the mentioned peptide T was digested with trypsin yielding a long (T-1) and a short (T-2) peptide

(Figure). Because of the unusual sequence (-Leu-Leu-Ile-Leu-) located at the carboxyl-terminal sequence of T-1, this peptide was cleaved at its 2 histidine residues with N-bromosuccinimide. Utilizing the resulting pentapeptide, Leu-Leu-Ile-Leu-Arg, the structure of the carboxyl-terminal region of CNBr-I could be established unambiguously.

The present study essentially completes the linear amino acid sequence of α_1 -acid glycoprotein. It should be noted that this is the first time that the amino acid sequence of a glycoprotein which possesses 5 carbohydrate units is described ^{14, 15}.

Zusammenfassung. Es gelang, die lineare Aminosäuresequenz von saurem α_1 -Glykoprotein (Orosomucoid), einem Globulin des menschlichen Plasmas, aufzuklären.

H. KAUFMANN and K. SCHMID

Department of Biochemistry, Boston University School of Medicine, Boston University Medical Center, 80 East Concord Street, Boston (Massachusetts 02118, USA), 19 August 1971.

- ¹ R. W. Jeanloz, in *Glycoproteins* (Ed. A. Gottschalk; B.B.A. Library, Elsevier Publishing Company, New York 1966), vol. 5, p. 362.
- ² P. V. Wagh, I. Bornstein and R. J. Winzler, J. biol. Chem. 244, 658 (1969).
- ³ H. Schachter, I. Jabbal, R. L. Hudgin and L. Pinteric, J. biol. Chem. 245, 1090 (1970).
- ⁴ K. Schmid, M. Ishiguro, J. Emura, S. Isemura, H. Kaufmann and T. Motoyama, Biochem. Biophys. Res. Commun. 42, 280 (1971).
- ⁵ T. IKENAKA, M. ISHIGURO, S. ISEMURA, H. KAUFMANN, W. BAUER and K. Schmid, Biochem. Biophys. Res. Commun. 42, 1142 (1971).
- ⁶ W. Bürgi and K. Schmid, J. biol. Chem. 236, 1066 (1961).
- ⁷ T. IKENAKA, H. BAMMERLIN and K. SCHMID, J. biol. Chem. 241, 5560 (1966).
- 8 М. Satake, Т. Окиуама, К. Ishihara and K. Schmid, Biochem. J. 95, 749 (1965).
- 9 R. W. Armentrout and R. F. Doolittle, Arch. Biochem. Biophys. 132, 80 (1969).
- T. GOLDBERGER and C. B. ANFINSEN, Biochemistry 1, 401 (1962).
 C. H. W. Hirs, Methods in Enzymology (Academic Press, New York
- 1967), vol. 11.

 12 S. IWANAGA, P. WALLEN, N. J. GROENDAHL, A. HENSCHEN and B. BLOMBÄCK, EUROP. J. Biochem. 8, 189 (1969).
- ¹³ M. Elzinga, C. T. Lai and C. H. W. Hirs, Arch. Biochem. Biophys. 123, 353 (1968).
- ¹⁴ This study was supported by grants from National Science Foundation No. GB-24813 and the National Institute of General Medical Sciences, U.S. Public Health Service No. GM-10374.
- ¹⁵ Acknowledgment. The authors are indepted to Dr. R. F. Doo-LITTLE, University of California, La Jolla, Calif., for his generous gift of lyophilyzed *Pseudomonas fluorescens* cells from which pyrrolidonecarboxylyl peptidase was isolated.